

METABOLIC & ENDOCRINE DISTURBANCES

Treatment of metabolic acidosis

in a haemodynamically stable baby:

where the base deficit ECF is >10 in a baby in whom there are no signs of hypovolaemia (such as increasing toe core gap, poor capillary refill) and no signs of hypotension;

where there is a base excess ECF of >10 the amount of bicarbonate which is given is a half correction which is calculated as follows:

$$0.3 \times \text{base excess} \times \text{baby's weight} \div 2$$

This is equivalent to the number of ml and half the number of millimoles of sodium bicarbonate that is required if 4.2% sodium bicarbonate is used (there is 0.5 mmol/ml of HCO_3 in 4.2%). This should be infused slowly at a rate of 1-2 ml/hr and no boluses of bicarbonate should be given over a period of <30 mins.

In small pre-term infants a figure of 0.4 is used because of the higher water content of the body (increased proportion of extra-cellular fluid).

1. Treatment of metabolic acidosis in a haemodynamically unstable baby:

Differential diagnosis of metabolic acidosis in the pre-term include:

- a) hypoxaemia or ischaemia secondary to perinatal asphyxia
- b) severe lung disease
- c) volume depletion
- d) myocardial dysfunction caused by immaturity, sepsis or asphyxia
- e) severe acidosis caused by neonatal metabolic disorder is rare but must be considered.

The most common causes in the haemodynamically unstable baby include hypovolaemia and hypotension.

If there are signs of hypovolaemia - such as increasing toe core gap, worsening metabolic acidosis, poor capillary refill - the baby will need to be corrected with the appropriate fluid volume, blood if there is any anaemia (see transfusion protocol), or significant natural or iatrogenic blood loss, and possibly fresh frozen plasma if there is a coagulation defect and in very rare cases either albumin or normal saline solutions. If there is hypotension and there is no sign of hypovolaemia, it is possible that you may need to use inotropes to counteract metabolic acidosis and hypotension (see hypotension protocol).

Inherited Metabolic Disorder

SAMPLES TO COLLECT WHEN AN INHERITED METABOLIC DISORDER IS SUSPECTED IN AN ACUTELY ILL BABY/CHILD

The most useful samples are those taken at the time of admission, when acutely ill, and this opportunity should not be missed. If a non-metabolic diagnosis is made subsequently, samples for the special biochemical tests can always be discarded.

1) Samples for essential preliminary tests

Haematology: Full blood count, differential and platelets
Clotting studies if suspect liver damage (bleeding, jaundice, hepatomegaly)

Biochemistry:

BLOOD Arterial blood gases and acid-base status; biochemical profile, including electrolytes and liver function tests, plasma glucose and plasma amino acids.

URINE Must be saved for organic acids and amino acids. The best sample is the first urine passed at the time of acute admission. Diagnostic abnormalities often disappear quickly following intravenous glucose or other resuscitation. A urine bag should be applied at one end, if possible, at least 5 ml of urine collected. This should be placed in a sterile urine bottle. Smell it! Test it (on the ward) with a Multistix, or similar (note especially, pH, ketones, glucose) and with a Clinitest. Then send it straight to the laboratory or, if out-of-hours, freeze it (ice cream compartment of ward specimen fridge). The next sample of urine passed should be saved as well.

Other tests at the time of presentation may be indicated by the clinical features, e.g. plasma ammonium (unexplained intractable fitting; Reye's syndrome-like illness); plasma insulin, growth hormone, cortisol (hypoglycaemia).

2) Hypoglycaemia without obvious cause - acute samples

(These do not apply to the majority of babies with 'common' neonatal hypoglycaemia!)

Blood for: glucose, insulin, growth hormone, cortisol, amino acids (with alanine quantification), 3-hydroxybutyrate, free fatty acids, liver function tests, lactate (if hepatomegaly). If possible, take 2 ml of 'spare' blood into a lithium heparin tube, which can be separated and stored for special tests (e.g. carnitine).

* The first urine passed must always be tested for ketones and stored for organic acid and amino acid analysis.

Urea and electrolytes and, perhaps, blood gas and acid-base status will be needed for management.

- and freeze urine.
2. Consider need for plasma lactate: ammonium.
 3. Take 2 or 3 ml of 'spare' plasma to be frozen, in case of need.
 4. Also take 5 ml blood into an EDTA tube, to be saved for future DNA analysis. The whole blood (not to be centrifuged!) may be stored at 4^o C and sent as soon as possible to the local tissue culture laboratory. If forewarned, they will advise about, or supply, appropriate medium.

In an emergency, skin biopsies may be kept overnight at 4^o C in a dry sterile container (tissue culture laboratories vary in their preference). **DO NOT FREEZE!**

Sterility is of paramount importance in collecting biopsies, particularly post-mortem. Skin should be cleansed thoroughly with, e.g., Hibitane and then dried with an alcoholic swab.

From a living baby - use local anaesthetic. The inner side of the forearm or the back of the arm, just above the elbow, are suitable sites.

After death - collect two biopsies from different sites and place into separate bottle of media.

5. If non-ketotic hyperglycinaemia is a possibility, 0.5-1 ml of CSF should be frozen (plain tube). If a congenital lactic acidosis, 0.5-1 ml CSF into a fluoride tube.
6. In rare circumstances in which deficiency of a liver enzyme which is not expressed in fibroblasts is a strong possibility (e.g. some urea cycle defect; von Gierke's disease) liver biopsy should be collected. If to be collected immediately post-mortem, it is kinder to obtain parental consent before death.

Two or three needle biopsies of tissue should be taken, wrapped in aluminium foil, and snap frozen in liquid nitrogen or solid carbon dioxide. Samples should be stored frozen, as cold as possible (-70^oC, or in liquid nitrogen preferably).

If histology, electron microscopy, or cytochemistry are required, additional tissue should be placed immediately into appropriate preservatives.

6) Special biochemistry tests

Samples to collect

- | | |
|----------------------------|--|
| * Plasma ammonium: | 2 ml (minimum) blood into EDTA tube
Must reach the laboratory within <u>10 mins</u> of collection |
| * Plasma lactate:
tube) | 2 ml (minimum) blood into fluoride tube (i.e. glucose
Must be taken straight to laboratory |
| Plasma amino acids: | 1 ml blood into lithium heparin tube |
| Plasma free fatty acids: | 1 ml into fluoride tube |

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Plasma for carnitine: 1 ml (minimum) into lithium heparin tube

CSF for glycine: 0.5-1ml into plasma tube

*CSF for lactate: 0.5-1ml into fluoride tube

Urine for organic and amino acids - at least 3 ml into a plain (MSSU) bottle. Send straight to laboratory during working hours. Freeze overnight if out-of-hours.
- if a dying child and very little urine, send whatever you can, however little!

Tissue culture fluid for skin biopsies - during working hours can be obtained from Histopathology, Level E, South Block.
- out-of-hours, put skin biopsies into a sterile (MSSU) bottle and keep at 4⁰C (**do not freeze**). Next day, contact the Cytogenetics laboratory at Guy's Hospital.

* These must only be sent after discussion with the laboratory, since they need immediate analysis.